MDCK Host Cell DNA Residue Detection Kit

Application: Quantitative analysis of DNA residue in recombinant protein expressed products, purified intermediate and finished products from host cell.

For Research, Development and Manufacturing Use Only!

Not for Therapeutic or Diagnostic Applications!

1. Introduction

This kit utilizes the TaqMan probe-based fluorescence quantitative PCR (qPCR) method. By employing highly specific primers and probes, it offers exceptional specificity and sensitivity, with a limit of quantification (LOQ) as low as 3 fg/μL. The kit is fully compatible with our Magnetic Residual DNA Sample Preparation Kit (Cat# CG-DP050 or CG-DP100), enabling seamless integration into your workflow.

The preparation process of DNA Control is completely consistent with that of the National Standard reference of host cell DNA y; therefore, it has high purity and no protein or ion interference. DNA Control has been calibrated by National Standard to ensure the accuracy of the sample quantitative detection.

The kit provides DNA Dilution Buffer, which enables good replicate parallelism in a single experiment and good reproducibility between multiple experiments.

2. Kit Components

DNA Amplification			
Components NO.	**Components Name**	**Cat#/Size**	
		CK-D050T(50T)	CK-D100T(100T)
B1	2XqPCR Mix	0.625mL	1.25mL
B2	Primer &Probe Mix	100μL	200μL
B3	DNA Dilution Buffer	2×1.5mL	4×1.5mL
B4	DNA Control (30ng/μL)	25μL	50μL
B5	RNase-Free H_2O	0.5mL	1mL
B6	50X ROX Reference Dye (Optional)	0.15mL	0.3mL

**The usage of ROX reference dye is optional and depends on the type of instrument being used. Please refer to the details in PART 6*

3. Equipment Required but Not Provided

1.	Pipette: 5μL-1000μL	**5.**	Mini Centrifuge
2.	1.5/2mL RNase-/DNase-free Centrifuge Tube	**6.**	DNase/RNase-free 8-Tube Strip
3.	200μL RNase-/DNase-free PCR Tube	**7.**	Biological Safety Cabinet Class 2
4.	Vortex Mixer	**8.**	Fluorescence qPCR Detection System

4. Shipping and Storage

1) All components are shipped with ice bags.
2) The kit should be stored at -20℃ and it is recommended to be used within one year. The component of B2 should be stored protected from light.
3) Components B2/B3/B4 can be stored at -20℃ for 2 years, while Components B1/B5/B6 can be stored at -20℃ for 1 year. B1/B5/B6 can also be purchased together as a separate set.

5. Preparation Before Experiment

1) Please read the manual thoroughly before use. All the components should be completely thawed, centrifuged at low speed, and vortexed well before use.
2) Avoiding the formation of bubbles, gently invert B1 (2XqPCR Mix) and B6 (50X ROX Reference Dye) separately after thawing them from -20℃ storage. Use the solution only after complete homogenization. **Note**: If component B1and B6 will be frequently used for a period of time, they can be stored at 2-8℃ for up to 3 months. Avoid repeated freeze-thaw cycles as much as possible. If not used after thawing, thoroughly mix before refreezing.
3) Do not pipette up and down repeatedly when mixing B2 (Primer & Probe Mix) and B4 (DNA Control) before use. Instead, a technique similar to cleaning the walls of the tube can be used to ensure the standard is evenly mixed. Note: To reduce the number of freeze-thaw cycles and avoid contamination, it is recommended to aliquot and store B4 (DNA Control) at -20℃ upon first use. (For detailed procedure, refer to the standard dilution protocol)
4) The thawed but unused B3 (DNA Dilution Buffer) can be stored at 2-8℃ for up to 7 days. If not used for an extended period, it should be stored at -20℃ to maintain stability.
5) For your safety and health, please wear lab coat and disposable gloves when conducting the experiment. UV irradiation for 30 minutes before and after the experiment is recommended to eliminate potential DNA contamination in the environment.
6) Due to the high sensitivity of fluorescence quantitative PCR experiments, it is very important to maintain a clean working environment. Before starting the experiment, it is recommended to thoroughly clean the pipette and the surrounding work area, and remove any unnecessary items during the experiment.

6. ASSAY PROCEDURE

1) Dilution of DNA Control and Preparation of Standard Curves

(1) Thaw B4 (DNA Control) and B3 (DNA Dilution Buffer) on ice, mix thoroughly by gentle vortexing, then centrifuge at low speed for 10 seconds.

(2) Prepare 7 clean 200μL PCR tubes, and label them as S0, S1, S2, S3, S4, S5 and S6. Add 45μL of DNA Dilution Buffer to each tube.

(3) Centrifuge the thawed DNA Control (30ng/μL) at high speed for 10 seconds, vortex for 5 seconds, then centrifuge it again for 10 seconds. (This DNA Control (30ng/μL) can be aliquoted and stored at -20°C for less than 3 months. Avoid repeated freeze-thaw cycles.)

(4) Add 5μL of the above homogenized DNA Control (30ng/μL) into a PCR tube labeled "S0" (non-standard curve concentration point). Centrifuge it at high speed for 10 seconds, vortex for 5 seconds, and centrifuge it again for 10 seconds. The resulting solution will have an intermediate concentration of 3000pg/μL, designated as S0.

(5) Dilution procedure in S1, S2, S3, S4, S5 and S6 tubes follows the same procedure as:

Tube	Dilution Procedure for the Standard Curve	Final Concentration
S1	5μL S0 + 45μL DNA Dilution Buffer	300pg/μL
S2	5μL S1 + 45μL DNA Dilution Buffer	30pg/μL
S3	5μL S2 + 45μL DNA Dilution Buffer	3pg/μL
S4	5μL S3 + 45μL DNA Dilution Buffer	300fg/μL
S5	5μL S4 + 45μL DNA Dilution Buffer	30fg/μL
S6	5μL S5 + 45μL DNA Dilution Buffer	3fg/μL

2) PCR Reaction System

Components	Volume(μL)
2X qPCR Mix	12.5
Primer & Probe Mix	2
DNA template (control or sample)	5
Add water	5.5
Total Volume	25

NOTES:

- Calculate the total volume of Mix solution required for this reaction based on the number of reaction wells: Mix solution = (number of reaction wells+4) * (12.5+2+5.5) μL (including the volume lost in the 4 wells). It is recommended to perform the operation on ice.
- Standards and samples are recommended to be tested in triplicate. The detection range of the standard curve mentioned above is suitable for most experiments and can be adjusted as needed, such as 30fg/μL to

300pg/μL.

- Maintain consistent experimental procedures. Seal the tube after sample addition, then centrifuge at low speed for 10 seconds to collect the liquid to the bottom. Vortex for at least 5 seconds to ensure complete mixing. Centrifuge again at low speed for 10 seconds. Remove the bubbles that may occurs.
- To ensure the accuracy of experimental results, we recommend diluting the protein concentration to 1-10mg/mL using 1X PBS for spike recovery experiments, which will ensure the recovery rate falls within the range of 50% to 150%.
- Matching concentration for ROX Reference Dye of several instruments are listed in the table below.:

Instrument	Concentration
ABI PRISM 7000/7300/7700/7900HT/Step One	2.5X（e.g. 1.25μL ROX/25μL system）
ABI 7500/7500Fast Stratagene Mx3000P/Mx3005P/Mx4000	0.5X（e.g. 0.25μL ROX/25μL system）
Roche/Bio-Rad/Eppendorf	No need to add

(1) Data Settings for PCR Instrument (Two-Step Method)

Phase	Temp.(℃)	Time	Content	Fluorescence Signal Collection	Cycles
Pre-denaturation	95℃	15mi	Pre-denaturation	NO	1
PCR Reaction	95℃	3sec	Denaturation *	NO	40
	60℃	30sec	Annealing/Extension *	YES	

NOTES:

- The pre-denaturation condition for PCR reaction must be set at 95℃ for 15 minutes.
- Select Reporter as "FAM", Quencher as "TAMRA".
- Follow the instrument user manual for time setting on different models. Set the time as 1 sec. for ABI 7900HT/7900HT Fast/ViiA 7/StepOnePlus.
- Annealing/Extension *: Follow the instrument user manual for time setting on different models. The time settings for several common instruments are listed in the table below:

Instrument	Time Setting
ABI 7500 Fast/7900HT/7900HT Fast/ViiA 7/StepOne/StepOnePlus	30 sec
Roche LightCycle/LightCycle 480	20 sec
ABI 7000/7300	31 sec
ABI 7500	32 sec

7. Criteria for Results

1) Standard Curve: $R^2 > 0.99$; Amplification Efficiency: 90%≤Eff%≤110%; Slope: -3.8~-3.1.

2) The recovery rate of spiked samples=(measured value of spiked samples - measured value of samples)/theoretical value of spiked samples * 100%, with a range of 50% -150%.

3) No Template Control (NTC): In the reaction system, replacing the target template with DNA Dilution Buffer while keeping other components unchanged, and the Ct value obtained should be 'Undetermined' or Ct value≥35.